

# Insect resistance

## Whitebacked planthopper populations on rice cultivars

K. S. Kushwaha, K. K. Mrig, and Rattan Singh, Haryana Agricultural University, Rice Research Station, Kaul-132021, Kurukshetra Haryana, India

Whitebacked planthopper (WBPH) *Sogatella furcifera* (Horvath) infestations occur throughout Haryana from the first week of September to harvest. Infested fields show hopperburn patches from late September.

Sixteen medium-duration (135-140 days) rice cultivars were evaluated during 1978 kharif for reaction to WBPH. They were grown in a randomized block design at 20- × 15cm spacing replicated 4 times. Jaya was included as a susceptible check.

Populations ranged from 17 to 222 nymphs/hill (see table). Variety RP79-8-3-2-1 had the lowest number of WBPH nymphs/hill, PR 106 had the highest. Of

Resistance of rice cultivars to whitebacked planthopper at Kaul, India.

Variety	Cross	Nymphs/hill <sup>a</sup>	
		x + 0.5	Av
RP979-583-2-1-1	RPA5981/Sona	4.17 a	17.075
PAU41-356-1-5	Phulpattas 72/mut. 65	5.79 b	33.15
RP975-284-2-2	Sona/RPW6-13	6.22 bc	38.475
RP6-516-33-6-1	TKM6/IR28	6.76 cd	45.275
Jaya	TNI/T141	6.79 cd	45.875
RP6-516-29-1	TKM6/IR8	6.98 d	48.375
RP6-1899-254	TKM6/IR8	7.17 de	51.225
Sona	GEB24/TN1	8.42 ef	70.725
HAU4-63-3	IR8/Jhona 349	8.73 f	76.025
PAU41-306-1-2	Phulpattas 72/mut. 65	8.80 f	77.25
RP6-516-34-1-8	TKM6/IR8	9.55 g	90.775
UPR70-30-42	IR8/Bas 370	9.74 g	94.475
PAU32-15-2	Bas 370/IR480-5	10.13 gh	102.25
RP633-519-1-3-4	IRI/KBJ-1//IR22	10.54 h	110.725
CR12-178	IR8/CR1014	13.69 i	187.167
PR106	IR8/Peta <sup>5</sup> //Bellepatna	14.89 i	221.625

<sup>a</sup>Av of 4 replications. Means followed by the same letter are not significantly different among themselves. Data transformed to x + 0.5.

16 cultivars tested, 9 developed hopperburn patches in one or more replications. Cultivars CR12-178 and PR 106 showed high susceptibility. Local cul-

tivar HAU4-63-3 was less susceptible than many of the cultivars tested. Resistant cultivars are now in minikit trials.

## Taxonomy of Asian and African rice gall midges

K. M. Harris, Commonwealth Institute of Entomology, London, UK, and R. J. Gagne, USDA Systematic Entomology Laboratory, Washington D.C., USA

It had been assumed until recently that the rice gall midge in Africa is the same species as the Asian rice gall midge *Orseolia oryzae* (Wood-Mason). Joint studies in 1981 by the Commonwealth Institute of Entomology and the USDA Systematic Entomology Laboratory of a

series of reared adults with associated larvae and pupae collected by Dr. J. Etienne, ISRA-Ziguinchor, from rice in Senegal show that the African rice gall midge is a morphologically distinct species. A formal description is being prepared.

## A technique for preparation of brown planthopper chromosomes

R. C. Saxena, associate entomologist, International Rice Research Institute, and senior research scientist, International Centre of Insect Physiology and Ecology, P. O. Box 30772, Nairobi, Kenya; and A. A. Barrion, graduate assistant, IRRI

Chromosome cytology has been used to determine subtle cytotoxic differences between related species, sibling species, subspecies, and biotypes. A simple and rapid technique for preparing meiotic chromosomes is needed to

examine and compare large insect samples, especially in studies of insect systematics and evolution. A technique developed at IRRI proved useful in preparing and studying brown planthopper chromosomes. The procedure has three steps.

1. Fixing and dissecting insects — Fifth-instar nymphs and newly emerged males collected from stock cultures at 0700 h are fixed in glass vials containing Carnoy's fluid — 1 part 99.7% glacial acetic acid and 3 parts 95% ethyl alcohol — for at least 2 minutes. A fixed insect is then dissected in a

drop of Ringer's solution on a clean glass slide. The head and thorax are discarded and the abdomen is dorsally incised to extract the tiny, translucent testes.

2. Staining, mounting, and labeling — The testes are submerged in a drop of 2% aceto-orcein or carmine solution for 2 minutes. With a fine-tipped, curved needle, each testis is macerated on a clean slide with a drop of 2% aceto-orcein. The cells are kept from drying out by adding a drop of 45% acetic acid. All debris are discarded.

A clean cover slip is placed over the